

REPORT DOCUMENTATION PAGE			Form Approved OMB No. 0704-0188	
Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing the collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden, to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Budget, Paperwork Reduction Project (0704-0188), Washington, DC 20503.				
1. AGENCY USE ONLY (Leave blank)		2. REPORT DATE 1/1/90-9/30/96		3. REPORT TYPE AND DATES COVERED Final Report
4. TITLE AND SUBTITLE Biological Applications of STM & AFM in Water/ High Resolution Microscopy of Nucleoprotein Complexes in Water			5. FUNDING NUMBERS G N00014-90-J-1455	
6. AUTHOR(S) S.M. Lindsay				
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) Arizona State University Tempe, Arizona 85287-1504			8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) Office of Naval Research 800 N. Quincy St Arlington, Virginia 22217-5660			10. SPONSORING / MONITORING AGENCY REPORT NUMBER	
11. SUPPLEMENTARY NOTES				
12a. DISTRIBUTION / AVAILABILITY STATEMENT Distribution Unlimited			12b. DISTRIBUTION CODE 19970122 140	
13. ABSTRACT (Maximum 200 words) Sample preparation methods and new instrumentation have been developed for the study of biological molecules in their native (aqueous) environment by scanning tunneling microscopy (STM) and atomic force microscopy (AFM). Information from STM is complicated by the many different electron transfer mechanisms, so structural information is difficult to obtain. However, chemical identification of certain electroactive molecules may be possible at the single molecule level. A new AFM with a magnetically-oscillated tip was used to generate images of DNA of unprecedented resolution. The DNA was imaged in-situ, spontaneously adsorbed to mica in the presence of divalent ions.				
14. SUBJECT TERMS STM/AFM, DNA, Biomolecular Structure, In-Situ Imaging			15. NUMBER OF PAGES 4	
			16. PRICE CODE	
17. SECURITY CLASSIFICATION OF REPORT U	18. SECURITY CLASSIFICATION OF THIS PAGE U	19. SECURITY CLASSIFICATION OF ABSTRACT U	20. LIMITATION OF ABSTRACT UL	

GENERAL INSTRUCTIONS FOR COMPLETING SF 298

The Report Documentation Page (RDP) is used in announcing and cataloging reports. It is important that this information be consistent with the rest of the report, particularly the cover and title page. Instructions for filling in each block of the form follow. It is important to **stay within the lines** to meet **optical scanning requirements**.

Block 1. Agency Use Only (Leave blank).

Block 2. Report Date. Full publication date including day, month, and year, if available (e.g. 1 Jan 88). Must cite at least the year.

Block 3. Type of Report and Dates Covered. State whether report is interim, final, etc. If applicable, enter inclusive report dates (e.g. 10 Jun 87 - 30 Jun 88).

Block 4. Title and Subtitle. A title is taken from the part of the report that provides the most meaningful and complete information. When a report is prepared in more than one volume, repeat the primary title, add volume number, and include subtitle for the specific volume. On classified documents enter the title classification in parentheses.

Block 5. Funding Numbers. To include contract and grant numbers; may include program element number(s), project number(s), task number(s), and work unit number(s). Use the following labels:

C - Contract	PR - Project
G - Grant	TA - Task
PE - Program Element	WU - Work Unit Accession No.

Block 6. Author(s). Name(s) of person(s) responsible for writing the report, performing the research, or credited with the content of the report. If editor or compiler, this should follow the name(s).

Block 7. Performing Organization Name(s) and Address(es). Self-explanatory.

Block 8. Performing Organization Report Number. Enter the unique alphanumeric report number(s) assigned by the organization performing the report.

Block 9. Sponsoring/Monitoring Agency Name(s) and Address(es). Self-explanatory.

Block 10. Sponsoring/Monitoring Agency Report Number. (If known)

Block 11. Supplementary Notes. Enter information not included elsewhere such as: Prepared in cooperation with...; Trans. of...; To be published in.... When a report is revised, include a statement whether the new report supersedes or supplements the older report.

Block 12a. Distribution/Availability Statement. Denotes public availability or limitations. Cite any availability to the public. Enter additional limitations or special markings in all capitals (e.g. NOFORN, REL, ITAR).

DOD - See DoDD 5230.24, "Distribution Statements on Technical Documents."

DOE - See authorities.

NASA - See Handbook NHB 2200.2.

NTIS - Leave blank.

Block 12b. Distribution Code.

DOD - Leave blank.

DOE - Enter DOE distribution categories from the Standard Distribution for Unclassified Scientific and Technical Reports.

NASA - Leave blank.

NTIS - Leave blank.

Block 13. Abstract. Include a brief (*Maximum 200 words*) factual summary of the most significant information contained in the report.

Block 14. Subject Terms. Keywords or phrases identifying major subjects in the report.

Block 15. Number of Pages. Enter the total number of pages.

Block 16. Price Code. Enter appropriate price code (*NTIS only*).

Blocks 17. - 19. Security Classifications. Self-explanatory. Enter U.S. Security Classification in accordance with U.S. Security Regulations (i.e., UNCLASSIFIED). If form contains classified information, stamp classification on the top and bottom of the page.

Block 20. Limitation of Abstract. This block must be completed to assign a limitation to the abstract. Enter either UL (unlimited) or SAR (same as report). An entry in this block is necessary if the abstract is to be limited. If blank, the abstract is assumed to be unlimited.

Final Report

GRANT #: N00014-90-J-1455

PRINCIPAL INVESTIGATOR: S.M. Lindsay

INSTITUTION: Arizona State University

GRANT TITLE: Biological Applications of STM and AFM in Water
/High Resolution Microscopy of Nucleoprotein Complexes in Water

AWARD PERIOD: (Jan. 1, 1990 - September 30, 1996)

OBJECTIVES: The work carried out under this grant was aimed at: (1) Developing reliable methods for imaging samples on surfaces under liquids and under potential control and (2) Applying these imaging techniques to problems in biomolecular structure.

ACCOMPLISHMENTS: The first goal was achieved, and, along the way, many contributions were made to the understanding of and technology of electrochemical scanning probe microscopy. The second goal was delayed as failures taught us much about the limitations of (and possibilities for) scanning tunneling microscopy applied to biological samples. The last year of the grant was spent applying methods developed for improving atomic force microscopy in 1992. The new technique (MA/C mode AFM) shows great promise for gentler imaging at higher resolution than heretofore possible.

CONCLUSIONS: (1) STM Images: A simple interpretation of the images is that they reflect *only the points where the molecule touches the metal substrate*. This information is adequate for, say following the path of a DNA molecule, but useless in most other cases. The limits of the technique are illustrated in some recent work in which we used covalent chemistry to bond DNA to gold substrates covalently. The oligomers used in that work gave understandable images and, subject to some assumptions, we even determined the bending angle in bulged DNA. In an (unpublished) study of ribozymes carried out in collaboration with F. Eckstein and T. Jovin of the Max Planck Institutes we showed that although the images were reproducible, 3D structural interpretation was not possible.

(2) AFM Imaging: Using MA/C mode, we have studied DNA microcircles spontaneously adsorbed onto mica in an aqueous solution [37]. The gentleness of the new mode is illustrated by our ability to obtain images in the presence of Mg alone. With tapping mode, Mg will not bind DNA spontaneously strongly enough to be imaged. We found a remarkable result: Axially strained microcircles had a small tendency to form kinks (localized sharp bends) in the presence of Mg. However, when the cation was changed to Zn, a four-fold increase in kinking occurred. Static bending was replaced by straight DNA connected by kinks (illustrated below). The same behavior is found in physiological solutions containing both Mg and Zn.

SIGNIFICANCE: The program has contributed significantly to what is now becoming routine use of the AFM for structural and micromechanical studies of biomolecules. The understanding of electron transfer in the STM has opened a new program aimed at electrochemical identification of single molecules.

PATENT INFORMATION:

"Method of Electrochemical Detection/Identification of Single Organic Molecules Using Scanning Tunneling Microscopy" Nongjian Tao and S.M. Lindsay US Patent 5,497,000, March 5, 1996.

"Controlled Force Microscope for Operation in Liquids" S.M. Lindsay US Patent 5,515,719, May 14, 1996.

AWARD INFORMATION: H. Willard Davis Lectureship in Chemistry, University of South Carolina (1994, Humbolt Senior Scientist Research Award (1993, Faculty Mentoring Award, ASU (1993), Vice-Chair Division of Biological Physics of the American Physical Society, 1994-5, Chair, Division of Biological Physics of the American Physical Society, 1995-6, Elected Fellow of the American Physical Society (1990) for: "*Pioneering studies in the application of scanning tunneling microscopy to imaging biomolecules, especially images of the DNA double helix in water*".

PUBLICATIONS:

- [1] S.M. Lindsay, O.F. Sankey, Y. Li and C. Herbst, J. Phys. Chem. **94**, 4655-4660 (1990).
- [2] T. Thundat, L.A. Nagahara, S.M. Lindsay and A. Majumdar, J. Vac. Sci. Technol. **B9**, 955-959 (1991).
- [3] Yu. L. Lyubchenko, S.M. Lindsay, J.A. DeRose and T. Thundat, J. Vac. Sci. Technol. **B9**, 1288-1290 (1991).
- [4] J.A. DeRose, S.M. Lindsay, L.A. Nagahara, P.I. Oden and T. Thundat, J. Vac. Sci. Technol. **B9**, 1166-1170 (1991).
- [5] S.M. Lindsay, Y. Li., J. Pan, T. Thundat, L.A. Nagahara, P.I. Oden, J.A. DeRose and U. Knipping, J. Vac. Sci. Technol. **B9**, 1096-1101 (1991).
- [6] P.I. Oden, T. Thundat, L.A. Nagahara, S.M. Lindsay, G.B. Adams and O.F. Sankey, Surface Science Letters **254**, L454-L459 (1991).
- [7] J.A. DeRose, T. Thundat, L.A. Nagahara and S.M. Lindsay, Surface Science **256**, 102-108 (1991).
- [8] Y. Li and S.M. Lindsay, Review of Scientific Instruments **62**, 2630-2633 (1991).
- [9] N.J. Tao and S.M. Lindsay, Applied Physics **70**, 5141-5143 (1991).
- [10] D.K. Luttrull, J. Graham, J.A. DeRose, D. Gust, T.A. Moore and S.M. Lindsay, Langmuir **8**, 765-768 (1992).
- [11] S.M. Lindsay, N.J. Tao, J.A. DeRose, P.I. Oden, Yu. L. Lyubchenko, R.E. Harrington and L. Shlyakhtenko, Biophysical Journal **61**, 1570-1584 (1992).
- [12] P.I. Oden, L.A. Nagahara, J. Graham, J. Pan, N.J. Tao, Y. Li, T.G. Thundat, J.A. DeRose and S.M. Lindsay, Ultramicroscopy **42-44**, 580-586 (1992).
- [13] N.J. Tao, J. Pan, Y. Li, P.I. Oden, J.A. DeRose, and S.M. Lindsay, Surface Science Letters **271**, L338-L344 (1992).
- [14] Y. Lyubchenko, L. Shlyakhtenko, R.E. Harrington, P.I. Oden and S.M. Lindsay, Proceedings of the National Academy of Science (USA) **90**, 2137-2140 (1993).
- [15] N.J. Tao and S.M. Lindsay, Surface Science Letters **274**, L546-L553 (1992).
- [16] N.J. Tao and S.M. Lindsay, Journal of Physical Chemistry **96**, 5213 - 5217 (1992).
- [17] N.J. Tao, S. M. Lindsay and S. Lees, Biophysical Journal, **63**, 1165-1169 (1992).
- [18] Y. Lyubchenko, B.L. Jacobs and S.M. Lindsay, Nucleic Acids Research **20**, 3983 - 3986 (1992).
- [19] A.A. Gall, L.S. Shlyakhtenko, R.E. Harrington, B.L. Jacobs, P.I. Oden and S.M. Lindsay, Journal of Biomolecular Structure and Dynamics **10**, 589-606 (1992).
- [20] N.J. Tao, J.A. DeRose and S.M. Lindsay, Journal of Physical Chemistry **97**, 910-919 (1993).
- [21] P.I. Oden, N.J. Tao and S.M. Lindsay, Journal of Vacuum Science and Technology, in press (1993).
- [22] Y.L. Lyubchenko, P.I. Oden, D. Lampner, S.M. Lindsay and K.A. Dunker, Nucleic Acids Research **21**, 1117-1123 (1993).
- [23] Y.Q. Li, N.J. Tao, J. Pan, A.A. Garcia and S.M. Lindsay, Langmuir, **9**, 637-641 (1993).

- [24] S.M. Lindsay, Y.L. Lyubchenko, N.J. Tao, Y.Q. Li, P.I. Oden and J. Pan, *Journal of Vacuum Science and Technology* **11**, 808-815 (1993).
- [25] J.A. DeRose, D.B. Lampner, S.M. Lindsay and N.J. Tao, *Journal of Vacuum Science and Technology* **11** 776-780 (1993).
- [26] T.W. Jing, A.M. Jeffrey, J.A. DeRose, Y.L. Lyubchenko, L.S. Shlyakhtenko, R.E. Harrington, E. Appella, J. Larsen, A. Vaught, D. Rekesh, F-X. Lu and S.M. Lindsay, *Proc. Natl. Acad. Sci. (USA)* **90**, 8934-8938 (1993).
- [27] A.M. Jeffrey, T.W. Jing, J.A. DeRose, A. Vaught, D. Rekesh, F-X. Lu and S.M. Lindsay, *Nucleic Acids Research* **21**, 5896-5900 (1993).
- [28] J. Pan, N.J. Tao and S.M. Lindsay, *Langmuir* **9**, 1556-1560 (1993).
- [29] S.M. Lindsay, T.W. Jing, A. Vaught and D. Rekesh, *Nanobiology*, **3**, 17-27 (1994).
- [30] J. Pan, T.W. Jing and S.M. Lindsay, *J. Phys. Chem.*, **98**, 4206-4208 (1994).
- [31] A. Vaught, T.W. Jing, S.M. Lindsay, *Chemical Physics Letters* **236** 306-310 (1995).
- [32] Th. Wandlowski, D. Lampner and S.M. Lindsay, *J. Electroanalytical Chemistry* **404**, 215-226 (1996).
- [33] Y.L. Lyubchenko, B.L. Jacobs, S.M. Lindsay and A. Stasiak, *Scanning Microscopy* **9**, 705-727 (1995).
- [34] D. Rekesh, Y. Lyubchenko, L.S. Shlyakhtenko and S.M. Lindsay, *Biophysical Journal* **71**, 1079 - 1086 (1996).
- [35] W. Han, S.M. Lindsay and T. Jing, *Applied Physics Letters* in press (1996).
- [36] Y.L. Lyubchenko, L.S. Shlyakhtenko, A. Nagaich, A. Appella, R.E. Harrington and S.M. Lindsay, *Scanning*, submitted (1996).
- [37] W. Han, M. Dlakic, R.E. Harrington and S.M. Lindsay, submitted to *Science*, 1996.
- [38] S.M. Lindsay in *Scanning Tunneling Microscopy: Theory, Techniques and Applications* (Ed. D. Bonnell, VCH Publishers) p 335-408 (1993).
- [39] S.M. Lindsay and M. Philipp, *Genetic Analysis* **8**, 8-13 (1991).
- [40] S.M. Lindsay and O.F. Sankey in *Scanned Probe Microscopies, STM and Beyond* ed K. Wickramasinghe, American Institute of Physics, NY, 125-135 (1992).
- [41] S.M. Lindsay, Y.L. Lyubchenko, A.A. Gall, L.S. Shlyakhtenko and R.E. Harrington, *SPIE proceedings of the international symposium on laser spectroscopy*, Los Angeles, January, 1992. pp 84-90.
- [42] S.M. Lindsay in *STM and SFM in Biology* M. Amrein and O. Marti (eds), Academic Press (1993).
- [43] S.M. Lindsay, J. Pan and T.W. Jing. *Proceedings of the Fall 1993 meeting of the Materials Research Society*, in press (1993).
- [44] S.M. Lindsay, T.W. Jing, J. Pan, D. Lampner, A. Vaught, J.P. Lewis and O.F. Sankey. *Proceedings of the NATO ASI on Nanoscale Probes of the Solid/Liquid interface*, eds. H. Seigenthaler and A.A. Gerwirth Kluwer, Netherlands, (1994) pp 25-43.